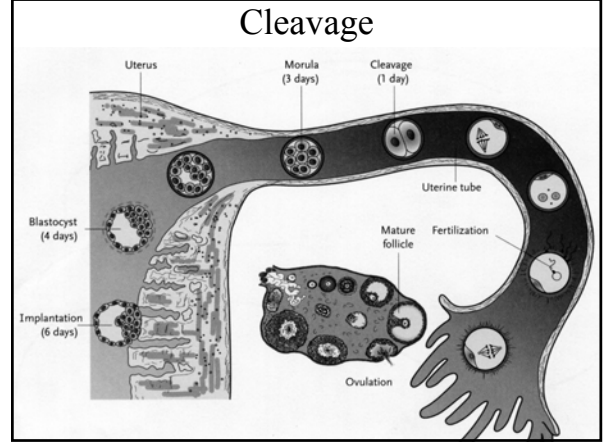


Cleavage

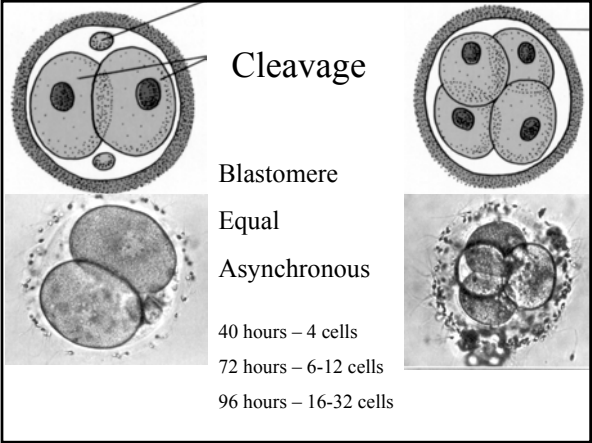
Cell Division – Cell Cycle Control
 Morula – Compaction
 Blastocyst – Hatching
 Implantation – Decidual Reaction
 Early Cell Lineages
 Inner Cell Mass
 Trophoblasts (Extra-embryonic)
 Anomalies



Cleavage – Molecular Events

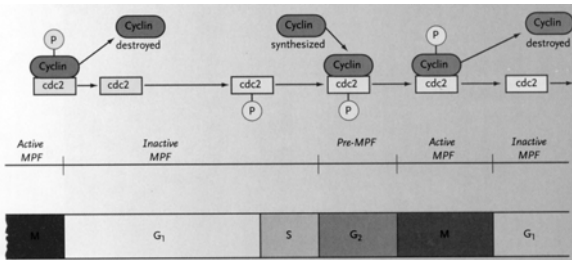
In mammals – no large maternal stores of RNA and ribosomes
 Zygotic transcription begins by 2-4 cell stage
Oct-3 – Transcription factor expressed in egg
 KO in mouse – arrest at 1 cell stage
 Expressed in blastomeres up to morula stage
 Expressed in germ cells

Cleavage



Blastomere
 Equal
 Asynchronous
 40 hours – 4 cells
 72 hours – 6-12 cells
 96 hours – 16-32 cells

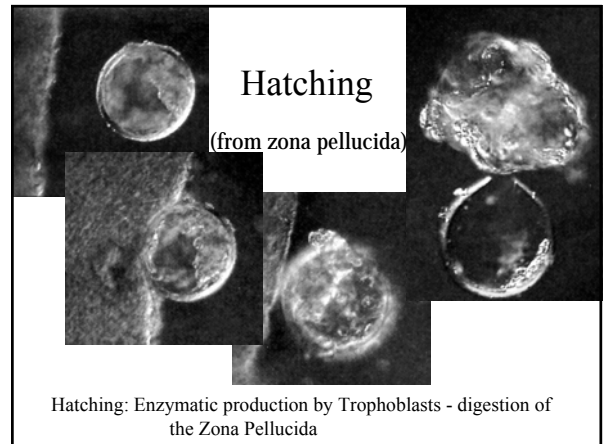
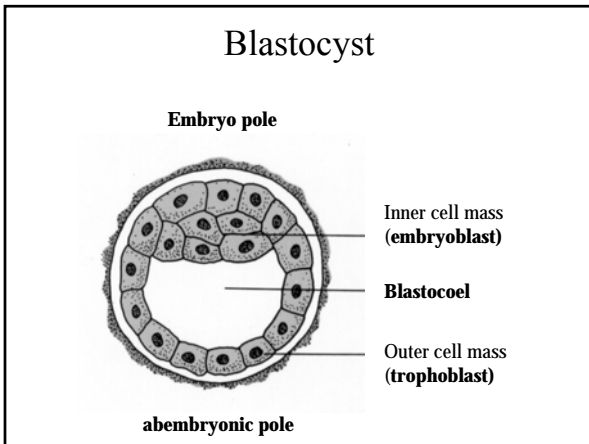
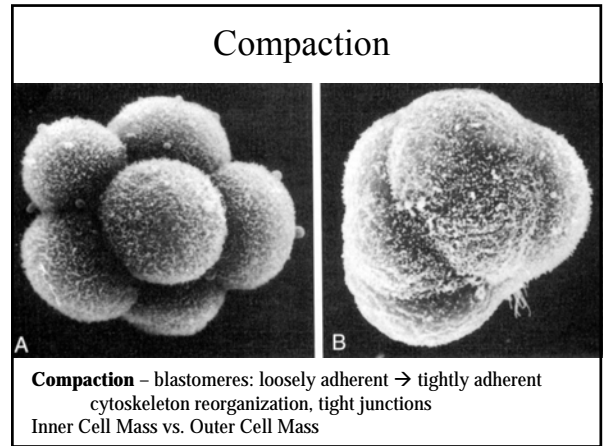
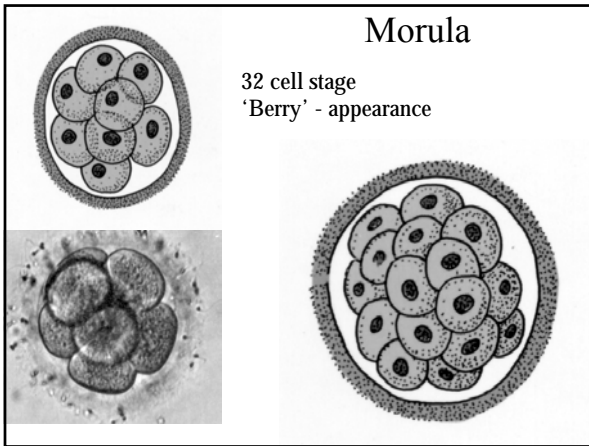
Cell Cycle Control



MFP = Maturation-promoting factor, or mitosis-promoting factor

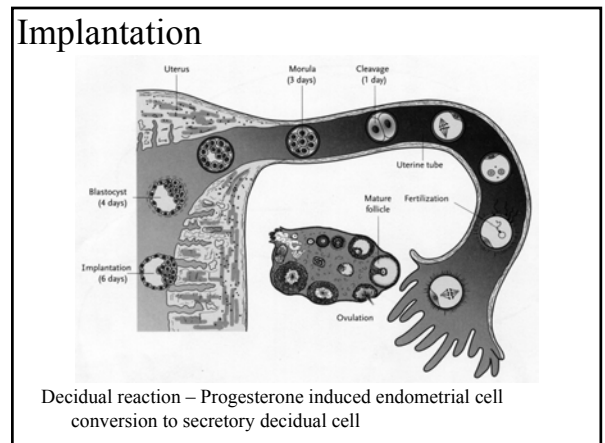
Cell Cycle Control

MPF – Mitosis Promoting Factor
 Heterodimer (cdc2 and cyclin B)
 Some Activities: Nuclear envelope breakdown, assembly of mitotic spindle
 Cdc2 – Cell Division Cycle 2
 Phosphoprotein (P in S and G2)
 Constitutively expressed
 Cyclin B – present in G2 and M
 Bound to cdc2
 Phosphoprotein (P in M)
 Degraded in G1



Zona Pellucida - Functions

- Species-specific sperm penetration
- Permanent block to polyspermy
- Acts as a porous selective filter - uterine tube signals
- Immunological barrier - no HLA (histocompatibility antigens)
- Keeps blastomeres together (loosely adherent)
- Prevents premature implantation



Implantation

Days 6-12

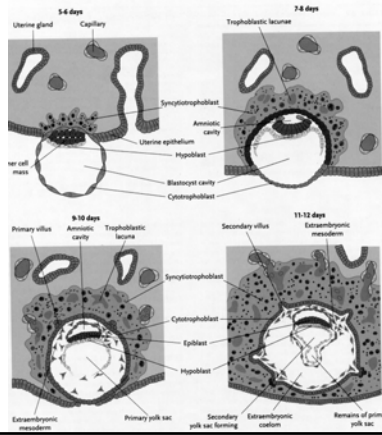
Adhesion, blastocyst to endometrium

Trophoblast proliferation

Syncytiotrophoblast

Secretion of hydrolytic enzymes

Breakdown of endometrium



Day 6

Blastocyst adheres to endometrium at embryo pole

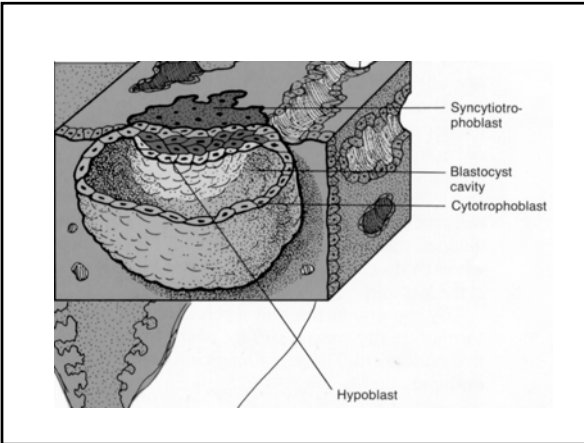
Trophoblast proliferation
production of hCG (maintains corpus luteum)

Embryo invasion

hydrolytic enzymes

Trophoblasts → Syncytiotrophoblast → hydrolytic enzymes

Trophoblasts → Cytotrophoblast



Day 7-8

Syncytiotrophoblast expansion

Lacunae form – filled with fluid (embryotroph)

Embryotroph provides nutrients to the embryo. Derived from maternal blood.

Embryo - Bilaminar germ disc:
Epiblast layer – cavitates to form the amniotic cavity.
Hypoblast layer form the exocoelomic cavity / primary yolk sac

Day 9-10

Lacunae enlarge

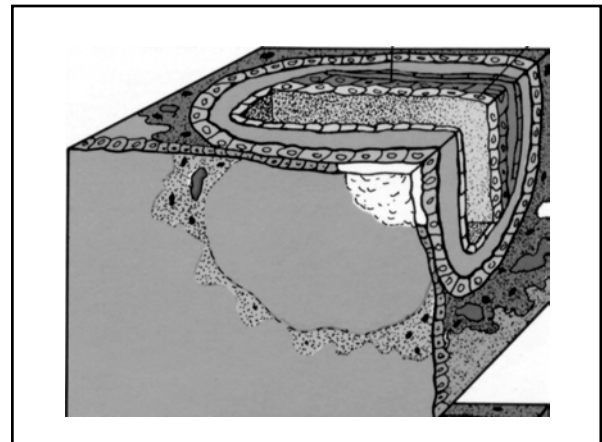
Syncytiotrophoblast expands around entire blastocyst

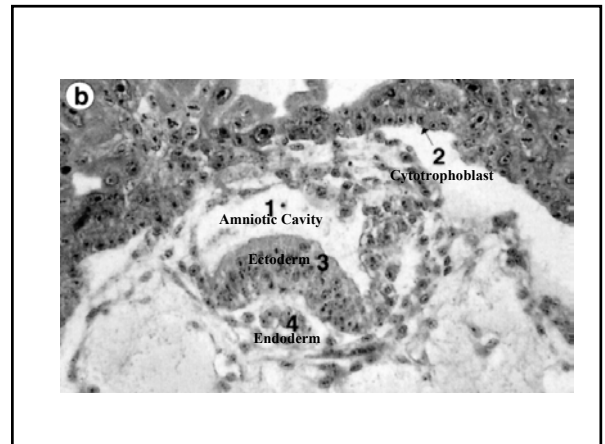
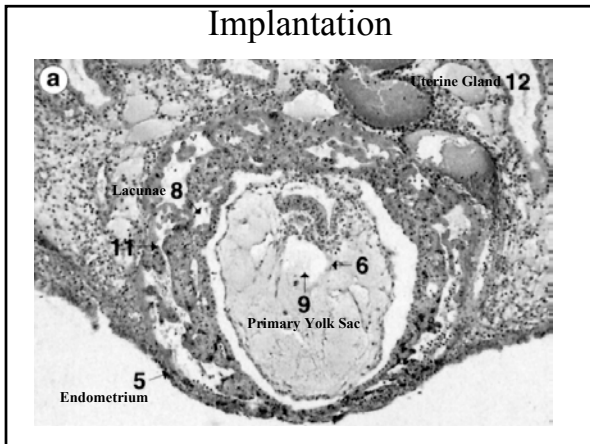
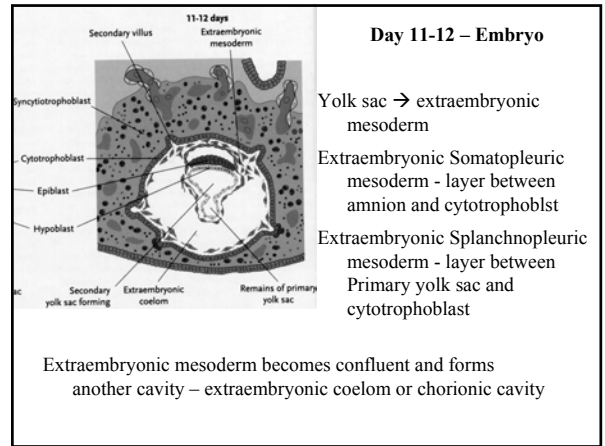
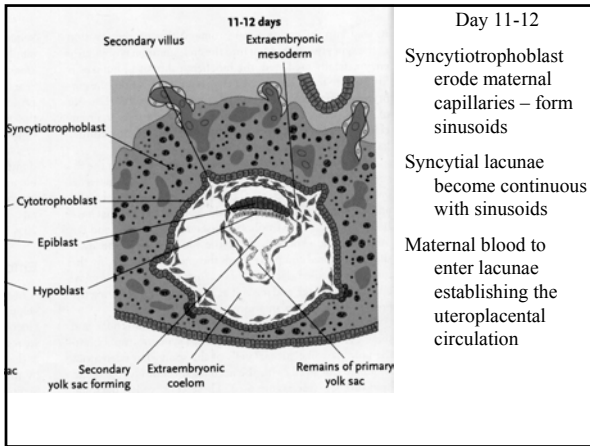
Cytotrophoblasts form primary villus – initiation of placenta formation

Implantation Complete

Coagulation Plug forms

Embryo: hypoblast → exocoelomic membrane = Hauser's membrane
Extraembryonic mesoderm from yolk sac





Summary

Day 0 Fertilization in Ampulla of uterine tube

Day 1 Zygotic transcription begins

Day 1-3 Cleavage – morula – compaction

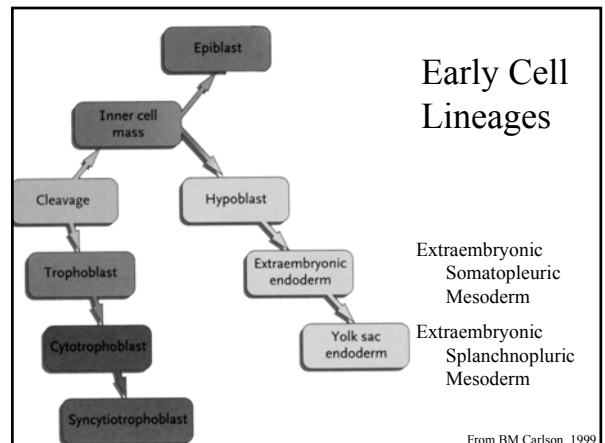
Day 3-4 Transport to uterine cavity
Relaxation of the uterotubal junction

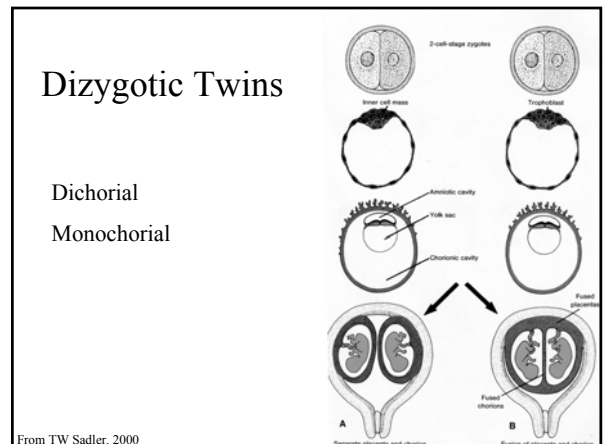
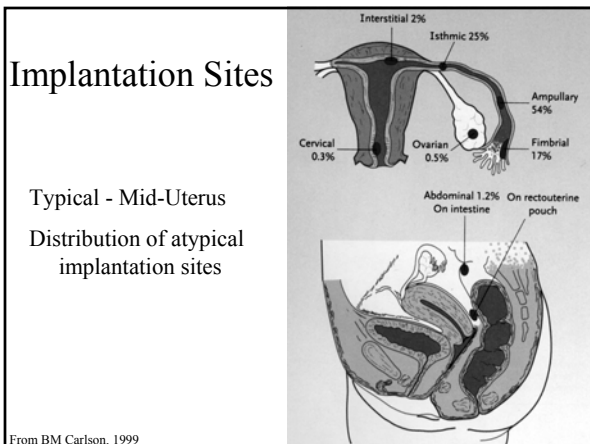
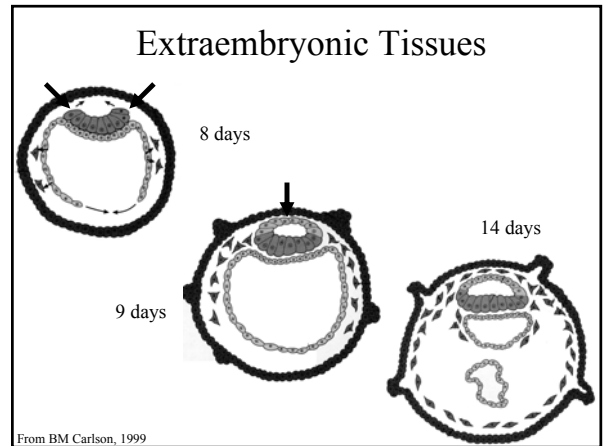
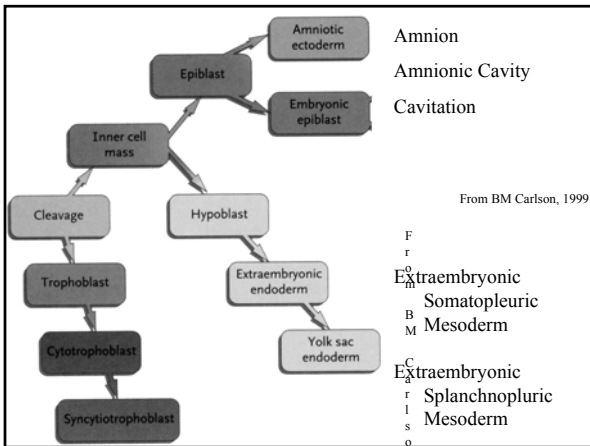
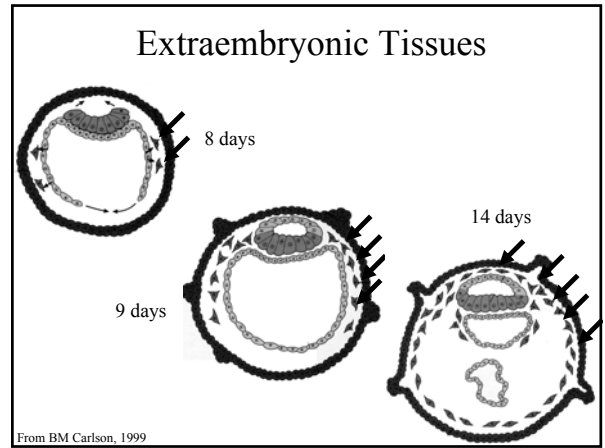
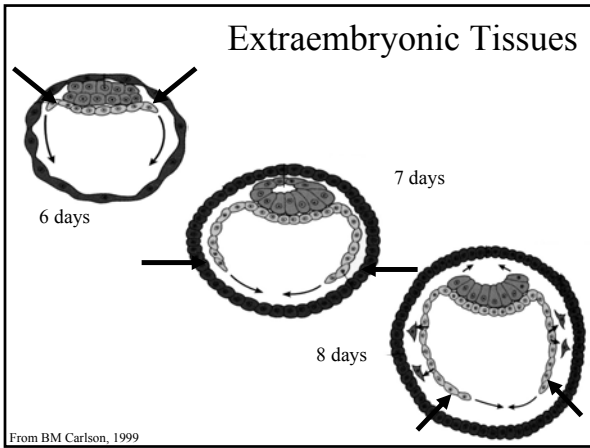
Day 5 Maturation of blastocyst, hatching

Day 6-7 Attachment / penetration of uterine stroma

Day 7-9 Invasion of uterine stroma

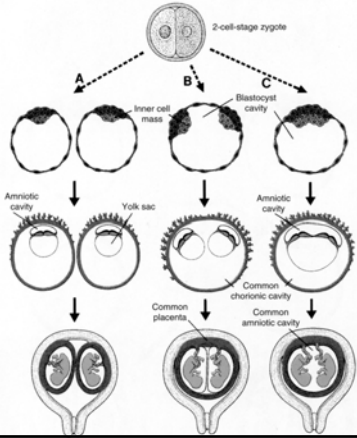
Day 9-11 Lacuna formation, erosion of spiral arteries



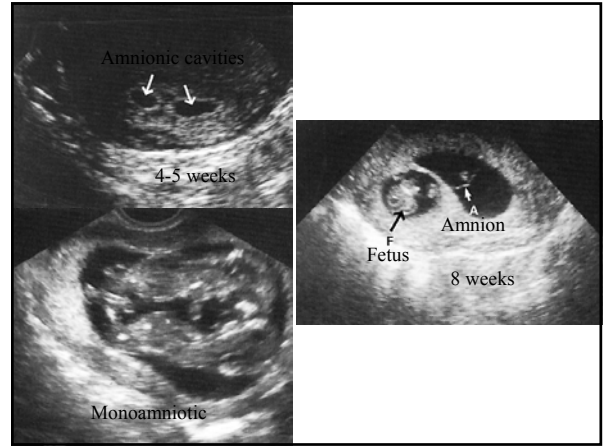


Monozygotic Twins

- Dichorial/Diamniotic (33%)
- Monochorial/Diamniotic (66%)
- Monochorial/Monoamniotic (rare)



From TW Sadler, 2000



Vanishing twins (triplets)
20% of twin pregnancies
Chromosomal or Structural abnormalities

Twins, Discordant Growth
Abdominal circumference, 3rd Trimester
>25% - associated with increased morbidity

Papyraceus – Death of a monozygotic co-twin
Circulatory interactions can cause problems

Table 16.1
Twinning rates per 1000 maternities by zygosity in different countries

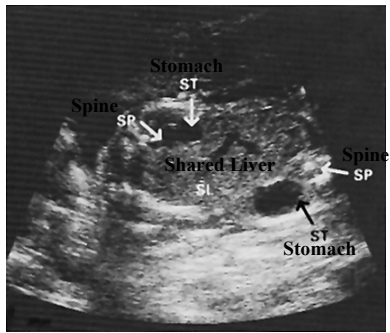
	Monozygotic	Dizygotic	Total
Nigeria	5.0	49.0	54.0
USA			
black	4.7	11.1	15.8
white	4.2	7.1	11.3
England and Wales	3.5	8.8	12.3
India	3.3	8.1	11.4
Japan	3.0	1.3	4.3

Table 16.2
Different monozygotic twin types

Time of division	Type of twinning
< 4 days	Dichorionic diamniotic
4–8 days	Monochorionic diamniotic
8–13 days	Monochorionic monoamniotic
> 13 days	Conjoined twins

Conjoined Twins

From BM Carlson, 1999



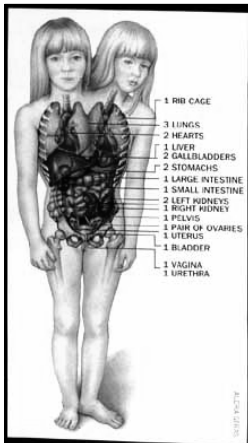
Ultrasound of conjoined twins



Daisy and Violet - English Siamese Twins

Pygopagus

Posterior union of the rump
19% of all conjoined twins.



Parapagus



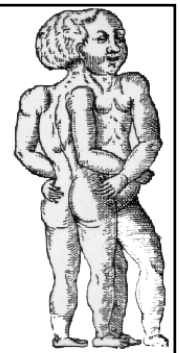
Three Months



Lateral union of the lower half



Cephalopagus



Anterior union of the upper half of the body with two faces on opposite sides of a conjoined head.
The heart is sometimes involved.

Cephalothoracopagus

Union of head and chest
There is only one brain
Hearts and gastrointestinal tracts are fused.

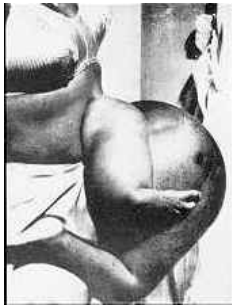


Craniopagus



Cranial fusion only

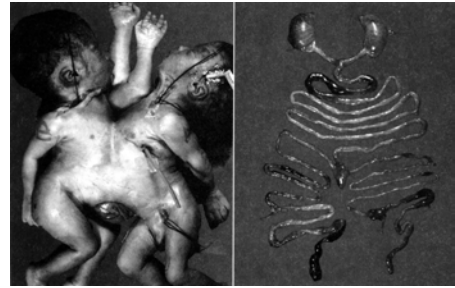
Parasitic Conjoined Twins



One twin without brain or heart

Thoracopagus

From BM Carlson, 1999



Anterior union of the upper half of the trunk.
The most common form of conjoined twins (about 75%)
Always sharing the heart.

Hydatidiform Mole

Pregnancy without an embryo (complete or partial mole)

Complete Mole = Only a placenta / No fetus – Diploid but with 2 sets of paternal chromosomes, no maternal contribution

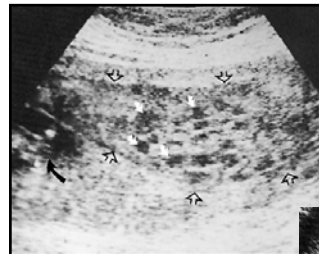
Partial Mole = Triploid (Maternal, 1N; Paternal, 2N)

Diagnosis – high hCG levels; ploidy analysis (flow cytometry)

1:1200 pregnancies in US; 1:200 pregnancies in Latin America/Asia

Hydatidiform Mole

Snow Storm appearance
Cystic Areas (white arrows)
No fetus
Placenta (open black arrows)



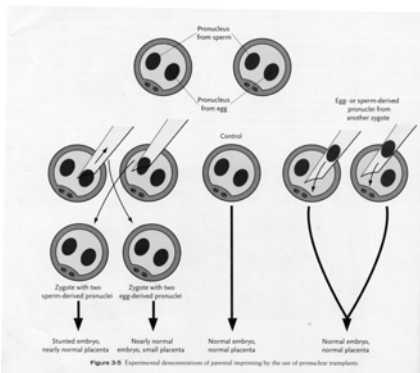
Partial Mole – cystic areas present

Fetus is present
commonly triploid
twin with mole in one sac
(rare)



From BM Carlson, 1999

Imprinting



Parental Imprinting

Identical genes derived from maternal and paternal DNA display differential expression
Selected genes are turned off during gametogenesis by methylation of certain bases
Imprinted patterns are not passed on to progeny, imprints erased during gametogenesis
Beckwith-Wiedemann syndrome - *Igf2*
Long arm Chr 15 deletion
Angelman's syndrome - Maternal deletion
Prader-Willi syndrome - Paternal deletion

Beckwith-Wiedemann Syndrome



Chromosome 11

- *Igf2* (Insulin-like Growth Factor) – growth promoter
- H19 – a growth suppressor

Mental deficiency – mild to moderate

Macrosomia – excessive growth, muscle, subcutaneous tissues

Macroglossia – protruding tongue, overgrowth of other craniofacial structures

Organ Hyperplasia – kidneys, pancreas

Angelman's Syndrome



“Happy Puppet Syndrome”

Maternal long arm of Chromosome 15 deletion

Severe mental deficiency – marked delays in motor milestones, absent speech, frequent laughter, frequent seizures

Puppet like gait

Widely spaced teeth

Macroglossia

Decreased ocular pigment → pale blue eyes

Prader-Willi Syndrome



Paternal long arm of Chromosome 15 deletion

Mental deficiency – mild-moderate

Normal birth size – decreased growth rate

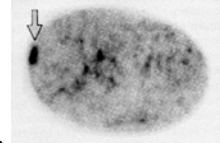
Short stature / Obesity

Very small hands, feet, genitalia

Fair skin, blue eyes, sun-sensitivity

Craniofacial – almond-shaped, narrow bifrontal diameter

X-Chromosome Inactivation



Inequality of Genetic Expression

Female-specific, 1 X-chromosome is inactive

Barr body – extreme condensation

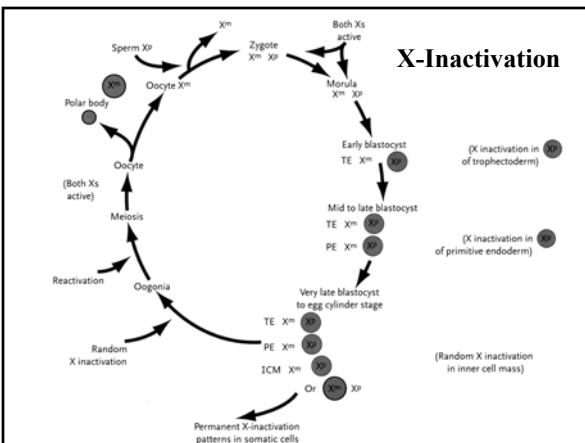
Both Xs are active thru cleavage

Blastocyst - Trophoblast – paternal X inactivated

Inner Cell Mass – both are active

Egg cylinder stage – differential X inactivation in cell lineages

Oogenesis – both Xs become active



Regulative Development

Ability of an embryo or organ to develop normally after removal or addition of parts

Fate of cells is not irreversibly fixed – influenced by environment

Contrast Mosaic Development

Fate Mapping studies

Developmental Potency – Totipotency

Stem Cells

